



Structure–activity relationship studies of sphingosine-1-phosphate receptor agonists with *N*-cinnamyl- β -alanine moiety

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ABSTRACT

Structure–activity relationship of sphingosine-1-phosphate receptor agonist was examined. In terms of reducing the flexibility of molecule, hit compound **1** was modified to improve S1P₁ agonistic activity as well as selectivity over S1P₃ agonistic activity. Novel S1P agonists with cinnamyl scaffold or 1,2,5,6-tetrahydropyridine scaffold were identified.

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Sphingosine-1-phosphate (S1P) exerts a variety of biological activities, including vascular maturation and cell survival.¹ Five S1P receptor subtypes have been known as S1P_{1–5}, respectively. One significant achievement in S1P research field was published in 2002. Lynch and co-workers reported that FTY720 (Fingolimod, Fig. 1), which was developed as a novel immunomodulator² and was recently approved in Russia and USA for the treatment of multiple sclerosis, is metabolized across species to a monophosphate ester, which can activate four S1P receptors (S1P_{1,3–5}) to sequester lymphocytes from circulation to secondary lymph tissue compartment.³ It is also reported that S1P₁ is essential for lymphocyte recirculation since S1P₁ modulates egress from thymus and peripheral lymphoid organs.⁴

It was reported that S1P₃ agonism does not relate to lymphocyte recirculation but links to bradycardia in rodents.⁵ Asymptomatic bradycardia was also reported in clinical studies of FTY720.⁶ Herein, we report our result to identify orally available S1P₁ agonists without S1P₃ agonism.

High Throughput Screening (HTS) campaign of our historical compound library yielded a hit compound **1**, which showed a sub-micromolar human S1P₁ agonistic activity with human S1P₃ and human S1P₅ agonistic activities. Additionally, oral administration of **1** in mice could induce peripheral lymphocyte lowering (PLL) with 50% effective dose (ED₅₀) of 26 mg/kg at 4 h after oral dosing (Fig. 2). Therefore, we decided to investigate the structure–activity

relationship around **1** to improve S1P₁ agonistic activity and selectivity against S1P₃.

Compounds in this report were synthesized in Schemes 1–3 as shown below. Compound **1** was prepared in 21% yield and compounds **9a–9c** were synthesized by using a solid-phase method⁷ from 4-(3-aminopropyl)phenol **2**. Starting from compound **2**, *N*-Boc- β -alanine derivative **5** was prepared in 56% yield. 2-Chlorotrityl resin was reacted with **5** and treated with the corresponding alcohol in the presence of tri-*n*-butylphosphine and *N,N,N,N*-tetramethylazodicarboxamide (TMAD). The treatment with the acetic acid and 2,2,2-trifluoroethanol yielded Boc-protected β -alanine **8a–8c**, which was treated with HCl/EtOAc to give **9a–9c** (Scheme 1).

Cinnamyl derivatives **12a–12d** were synthesized by reductive alkylation of β -alanine with aldehydes⁸ **11a–11d** in acceptable yields. Cinnamyl aldehyde **11a** was derived by reduction of the corresponding ester by LiAlH₄ in presence of Et₂NH⁹ in 17% yield. (4-Hydroxy)-acetophenone **13c** was treated with diethyl (cyanomethyl)-phosphonate and the resulting α,β -unsaturated nitrile was converted to the corresponding aldehyde, which was treated with alkyl bromide and each regioisomer generated under the reaction condition could be separated by column chromatography. The

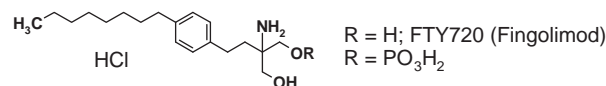


Figure 1. Structure of FTY720 (Fingolimod).

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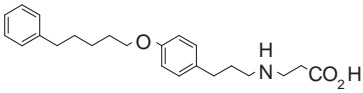
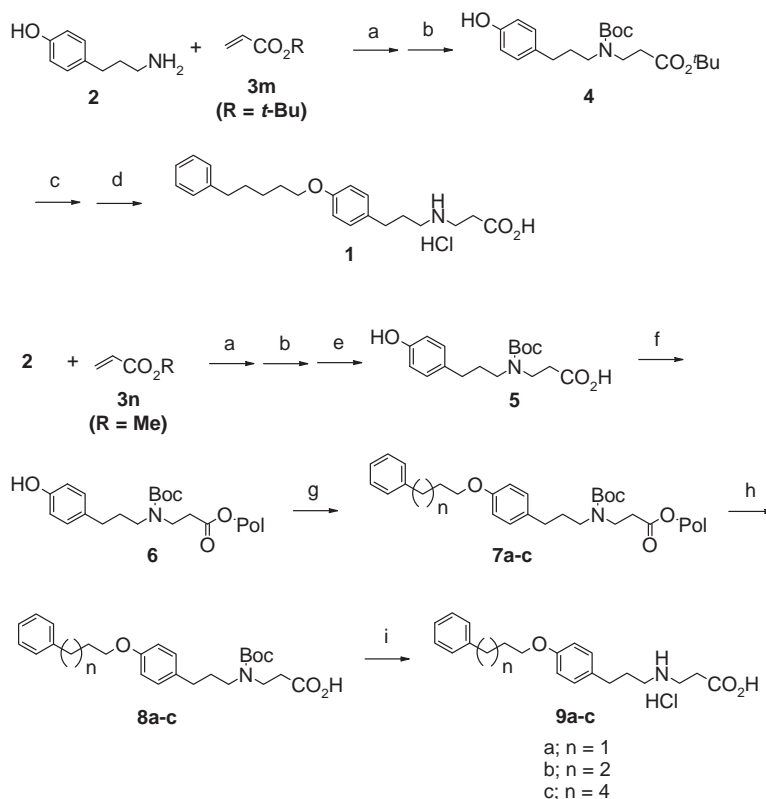
| Hit compound 1 | Ca Assay EC ₅₀ (uM) | | | | Mouse PLL ED ₅₀ (mg/kg) |
|---|-----------------------------------|-------------------|-------------------|-------------------|---------------------------------------|
| | hS1P ₁ | hS1P ₂ | hS1P ₃ | hS1P ₅ | <i>p.o.</i> 4 hr |
|  | 0.16 | >10.0 | 4.2 | 0.17 | 26 |

Figure 2. Structure and pharmacological profiles of hit compound 1.



Scheme 1. Reagent: (a) MeOH; (b) Boc₂O, THF; (c) Ph(CH₂)₅Br, K₂CO₃, DMF; (d) HCl/1,4-dioxane; (e) NaOH-aq, MeOH, THF; (f) Pol-Cl (Pol = 2-chlorotriptyl resin), *i*-Pr₂NEt, CH₂Cl₂; (g) Ph-(CH₂)_n-CH₂CH₂OH, *n*-Bu₃P, TMAD, CH₂Cl₂, THF; (h) CF₃CH₂OH, AcOH, CH₂Cl₂; (i) HCl/EtOAc.

separated *trans*- and *cis*- α,β -unsaturated aldehydes **11c** and **11d** were reacted with β -alanine to yield **12c** and **12d**, respectively (Scheme 2).

Tetrahydropyridine derivative **18** and piperidine derivative **19** were prepared via tetrahydro pyridine derivative **17**, which was obtained from the corresponding piperidone **15**. Piperidone **15** was reacted with lithiated phenylbromide **14** and then converted to tetrahydro pyridine **17** by HCl deprotection of Boc group followed by dehydration and Michael addition to *tert*-butyl acrylate in 52% yield. Tetrahydropyridine derivative **17** was converted to **18** by the treatment with trifluoroacetic acid (TFA) in 82% yield, and **19** by hydrogenation and subsequent treatment with TFA in 52% yield (Scheme 3).

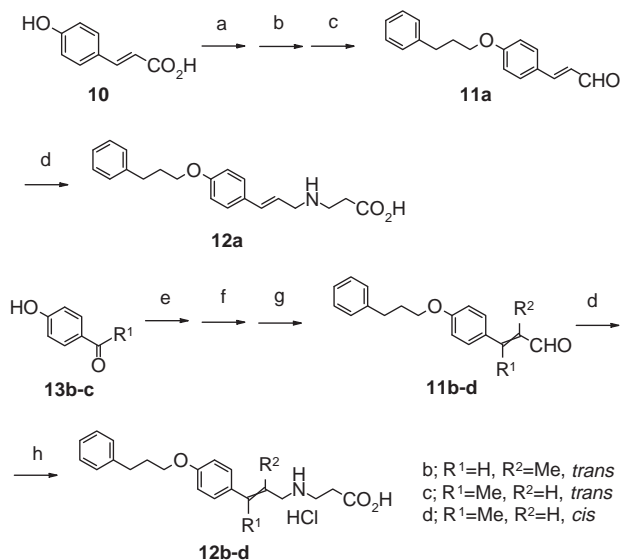
Piperidine derivative **24** was prepared from 3-hydroxypiperidine **20**. Piperidine derivative **20** was alkylated and oxidised to give **21** in 77% yield. On the other hand, phosphonium salt **23** was obtained from benzyl alcohol **22** in 94% yield. Phosphonium salt **23** was treated with dimsyl anion to afford a phosphonium ylide and reacted with ketone **21** to yield the condensed product in regioisomeric mixture, which was deprotected by HCl and then separated by column chromatography to give the *trans* isomer **24** (Scheme 3).

The effect of the length of alkylene linker between terminal phenyl and internal phenyl ring of hit compound **1** on its *in vitro*

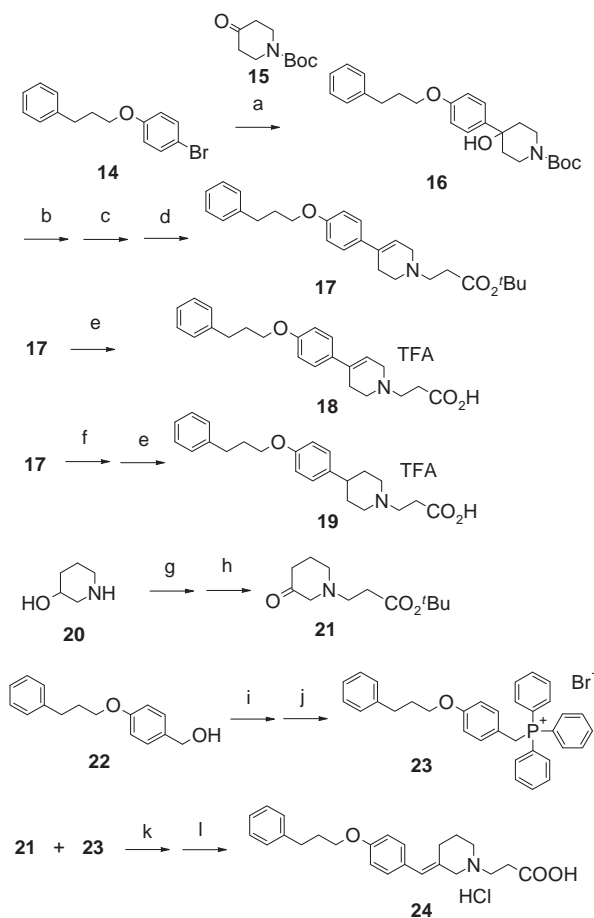
activity and oral exposure level was investigated. Although compound **1** showed the highest activity against S1P₁, shorter analogues **9a** and **9b** as well as longer analogue **9c** retained the S1P₁ agonistic activity. On the other hand, compound **9a** showed higher oral exposure level in rat than compound **1** (Table 1), which supports that the restriction of conformational flexibility improves the oral absorption.¹⁰ We decided to use propylene linker for the further SAR studies.

Next we modified the propylene linker between the internal phenyl ring and nitrogen (NH) in compound **9a** by introducing a double bond. Cinnamylamine compound **12a** significantly improved not only S1P₁ agonist activity but also selectivity over S1P₃. Hale et al. reported the S1P₁ selective agonists with benzyl amine moiety at the γ -position of carboxylic acid.¹¹ By combining this information, it is supposed that the π -electron density at this position might be important for S1P₁ agonist activity (Table 2).

Introducing a methyl group on the double bond of the allyl group of **12a** was examined. Whereas compound **12b** with methyl group at the β -position of amine showed comparable S1P₁ agonistic activity and selectivity over S1P₃, compound **12c** with methyl group at the γ -position improved both S1P₁ activity and selectivity. Additionally, the activity to reduce the peripheral blood lymphocyte count was enhanced significantly. Compound **12d** with *cis*-double bond showed less S1P₁ agonistic activity than



Scheme 2. Reagent: (a) SOCl₂, MeOH; (b) Ph(CH₂)₃OH, Ph₃P, DEAD, THF; (c) LiAlH₄, Et₂NH, *n*-hexane; (d) (i) H₂NCH₂CH₂CO₂H, NaOH, HC(OMe)₃, MeOH, THF; (ii) NaBH₄; (e) (EtO)₂P(O)CHR²CN (b: R² = Me, c: R² = H), NaH, THF; (f) DIBAL, THF; (g) Ph(CH₂)₃Br, K₂CO₃, DMF; (ii) separation of regioisomer **11c** and **11d**; (h) HCl-aq/THF.



Scheme 3. Reagent: (a) *n*-BuLi, THF; (b) HCl/1,4-dioxane, CH₂Cl₂; (c) Et₃SiH, TFA, CH₂Cl₂; (d) CH₂=CHCO₂Bu^t, *i*Pr₂NEt, DMF; (e) TFA, CH₂Cl₂; (f) H₂, 10% Pd-C, EtOAc; (g) CH₂=CHCO₂Bu^t, MeOH; (h) (COCl)₂, DMSO, Et₃N, CH₂Cl₂; (i) CBr₄, Ph₃P, CH₂Cl₂; (j) Ph₃P, PhMe; (k) NaH, DMSO; (l) (i) HCl/EtOAc, (ii) separation of regioisomers.

Table 1

The influence of length of linker towards S1P₁ and S1P₃ agonistic activities and oral exposure levels in rat

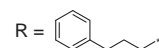
| Compound | <i>n</i> | Ca assay ^a EC ₅₀ (μM) | | Pharmacokinetic properties ^b in rat (30 mg/kg p.o.) | | |
|-----------|----------|---|-------------------|--|------------------------|--------------------|
| | | hS1P ₁ | hS1P ₃ | AUC _{inf} μg h/mL | C _{max} μg/mL | T _{1/2} h |
| 9a | 1 | 2.6 | 9.0 | 174 (±22) | 28 (±6.4) | 3.8 (±0.4) |
| 9b | 2 | 1.5 | >10 | N.T. | N.T. | N.T. |
| 1 | 3 | 0.16 | 4.2 | 11 (±1.1) | 2.1 (±0.23) | 3.8 (±0.1) |
| 9c | 4 | 2.7 | >10 | N.T. | N.T. | N.T. |

^a Agonistic activity was evaluated by the measurement of intracellular calcium concentration stimulated by the addition of each compound for human S1P₁ or human S1P₃ receptors stably expressed in Chinese Hamster Ovary (CHO) cell, respectively.

^b Values are means of three experiments, standard deviation is given in parentheses (N.T. = not tested).

Table 2

Replacement of propylene linker to allyl linker



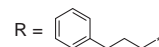
| Compound | Structure | Ca assay ^a EC ₅₀ (μM) | | Mouse PLL ^b ED ₅₀ (mg/kg) p.o. 4 h |
|------------|-----------|---|-------------------|--|
| | | hS1P ₁ | hS1P ₃ | |
| 9a | | 2.6 | 9.0 | N.T. |
| 12a | | 0.070 | 3.3 | 4.6 |

^a See Table 1.

^b Individual data points for dose-titrations were the average percentage decrease of peripheral blood lymphocyte counts in *n* = 5 animals versus control (*n* = 5) 4 h after oral administration of the test compound (N.T. = not tested).

Table 3

Addition of a methyl group on double bond



| Compound | Structure | Ca assay ^a EC ₅₀ (μM) | | Mouse PLL ^b ED ₅₀ (mg/kg) p.o. 4 h |
|------------|-----------|---|-------------------|--|
| | | hS1P ₁ | hS1P ₃ | |
| 12b | | 0.033 | 1.9 | N.T. |
| 12c | | 0.021 | >10.0 | 0.69 |
| 12d | | 0.93 | >10.0 | N.T. |

^{a,b} See Tables 1 and 2.

trans-congener **12c** (Table 3), which indicates the *trans* configuration might be beneficial to assist the basic nitrogen for interacting with S1P₁ receptor. This unexpected result prompted us to investigate further active conformations of compound **12a**.

Compound **12a** should have two distinct conformations around the allyl amine linker: extended conformer A and folded conformer

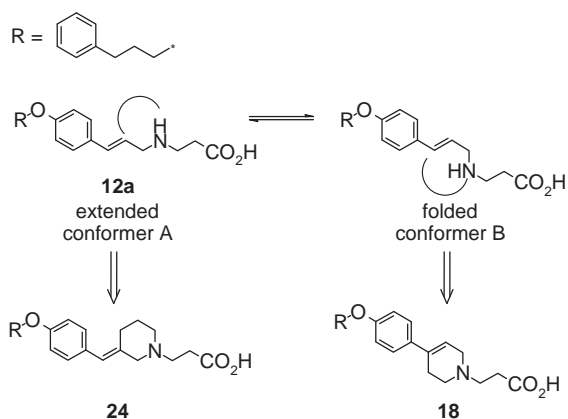


Figure 3. Conformational relationship between compounds **18** and **24**.

Table 4
Cyclic derivatives of compound **12a**

| Compound | Structure | Ca assay ^a EC ₅₀ (μ M) | | Mouse PLL ^b ED ₅₀ (mg/kg) p.o. 4 h |
|-----------|-----------|--|-------------------|--|
| | | hS1P ₁ | hS1P ₃ | |
| 18 | | 0.045 | 1.2 | 1.4 |
| 24 | | 1.5 | >10.0 | N.T. |
| 19 | | 1.6 | >10.0 | N.T. |

^{a,b} See Tables 1 and 2.

B (Fig. 3). Compounds **24** and **18** were designed to fix the extended conformation A and folded conformer B, respectively.

As a result, compound **18** showed much stronger S1P₁ activity than compound **24** (Table 4). This data strongly suggests that the folded conformer B would be an active conformation of compound **12a**. Additionally, the activity of compound **19**, having a single bond instead of double bond in the piperidine ring compared to compound **18**, was weaker than compound **18**, which indicates again that the π -electron density at this position might be important to activate S1P₁ receptor.

In summary, we explored SAR of hit compound **1** to identify a cinnamyl derivative **12a** with improved in vitro S1P₁ agonist activity as well as selectivity over S1P₃. Further modification of compound **12a** led to compound **12c** with strong S1P₁ agonism

and selectivity over S1P₃. Compound **12c** also showed superior pharmacokinetic property with long half-time ($T_{1/2}$ = 7.4 h) in rat (data not shown). During the SAR study, the importance of π -electron density between the central benzene ring and nitrogen atom was also determined. Furthermore, we conclude that the active conformation of compound **12a** would be the folded conformer **18**. Further optimisation of compounds **12c** and **18** will be reported in due course.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2011.01.029. These data include MOL files and InChIKeys of the most important compounds described in this article.

References and notes

- (a) Lee, M.-J.; Thangada, S.; Claffey, K. P.; Ancellin, N.; Liu, C. H.; Kluk, M.; Volpi, M.; Shâafi, R. I.; Hla, T. *Cell* **1999**, *99*, 301; (b) Cuvillier, O.; Pirianov, G.; Kleuser, B.; Vanek, P. G.; Coso, O. A.; Gutkind, J. S.; Spiegel, S. *Nature* **1996**, *381*, 800; (c) Marsolais, D.; Rosen, H. *Nat. Rev.* **2009**, *8*, 297.
- Fujita, T.; Hirose, R.; Yoneta, M.; Sasaki, S.; Inoue, K.; Kiuchi, M.; Hirase, S.; Chiba, K.; Sakamoto, H.; Arita, M. *J. Med. Chem.* **1996**, *39*, 4451.
- Brinkmann, V.; Davis, M. D.; Heise, C. E.; Albert, R.; Cottens, S.; Hof, R.; Bruns, C.; Prieschl, E.; Baumruker, T.; Hiestand, P.; Foster, C. A.; Zollinger, M.; Lynch, K. R. *J. Biol. Chem.* **2002**, *277*(24), 21453.
- Matloubian, M.; Lo, C. G.; Cinamon, G.; Lesneski, M. J.; Xu, Y.; Brinkmann, V.; Allende, M. L.; Proia, R. L.; Cyster, J. G. *Nature* **2004**, *427*, 355.
- (a) Forrest, M.; Sun, S.-Y.; Hajdu, R.; Bergstrom, J.; Card, D.; Doherty, G.; Hale, J.; Keohane, C.; Meyers, C.; Milligan, J.; Mills, S.; Nomura, N.; Rosen, H.; Rosenbach, M.; Shei, G.-J.; Singer, I. I.; Tian, M.; West, S.; White, V.; Xie, J.; Proia, R. L.; Mandala, S. *J. Pharmacol. Exp. Ther.* **2004**, *309*, 758; (b) Himmel, H. M.; Heringdorf, D. M. Z.; Graf, E.; Dobrev, D.; Kortner, A.; Schüller, S.; Jakobs, K. H.; Ravens, U. *Mol. Pharmacol.* **2000**, *58*, 449; (c) Koyrakh, L.; Roman, M. I.; Brinkmann, V.; Wickman, K. *Am. J. Transplant.* **2005**, *5*, 529; (d) Sanna, M. G.; Liao, J.; Jo, E.; Alfonso, C.; Ahn, M.-Y.; Peterson, M. S.; Webb, B.; Lefebvre, S.; Chun, J.; Gray, N.; Rosen, H. *J. Biol. Chem.* **2004**, *279*, 13839.
- (a) Tedesco-Silva, H.; Mourad, G.; Kahan, B. D.; Boira, J. G.; Weimar, W.; Mulgaonkar, S.; Nashan, B.; Madsen, S.; Charpentier, B.; Pellet, P.; Vanrenterghem, Y. *Transplantation* **2004**, *77*, 1826; (b) Budde, K.; Schmouder, R. L.; Brunkhorst, R.; Nashan, B.; Lückner, P. W.; Mayer, T.; Choudhury, S.; Skerjanec, A.; Kraus, G.; Neumayer, H. H. *J. Am. Soc. Nephrol.* **2002**, *13*, 1073.
- (a) Mellor, S. L.; McGuire, C.; Chan, W. C. *Tetrahedron Lett.* **1997**, *38*, 3311; (b) Barlos, K.; Gatos, D.; Kallitsis, J.; Papaphotiu, G.; Sotiriou, P.; Wenqing, Y.; Schafer, W. *Tetrahedron Lett.* **1989**, *30*, 3943.
- Verardo, G.; Geatti, P.; Pol, E.; Giumanini, A. G. *Can. J. Chem.* **2002**, *80*, 779.
- Zweifel, G.; Hahn, G. R.; Shoup, T. M. *J. Org. Chem.* **1987**, *52*, 5486.
- Veber, D. F.; Johnson, S. R.; Cheng, H.-Y.; Smith, B. R.; Ward, K. W.; Kopple, K. D. *J. Med. Chem.* **2002**, *45*, 2615.
- Hale, J. J.; Doherty, G.; Toth, L.; Li, Z.; Mills, S. G.; Hajdu, R.; Keohane, C. A.; Rosenbach, M. J.; Milligan, J.; Shei, G.-J.; Chrebet, G.; Bergstrom, J.; Card, D.; Rosen, H.; Mandala, S. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 3495.